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# Headspace Techniques for Flavor Characterization and Off-Odor Detection within the Food Industry

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# **INTRODUCTION**

More often than not the difference of odors and flavors in the same product is due to very small concentrations of specific chemicals. Potency of the odor is a function of both concentration of odor causing chemicals in the sample and their affinity to the sample matrix (partition coefficient). Any modification of matrix such as digestion, addition of solvents or salts may alter the composition of the headspace as compared with the original odor. It is desirable to characterize the odors/flavors over condensed matrices without any sample preparation or any typical analytical manipulations used to enhance the headspace concentration. The headspace conditions should not be altered from the conditions of human sensory testing.

## **DISCUSSION**

Analysis of a single aliquot of the headspace (regular headspace sampling) is often used to determine the profile of a consumer food product but is typically insufficient to provide the necessary sensitivity to determine the cause of a "bad" taste or odor. This type of trace detection work requires much larger volumes of headspace and an ability to pre-concentrate these contaminants prior to GC analysis. The Markelov HS9000 Static and Dynamic Headspace system utilizes a patented sampling needle which is comprised of two passages as shown in *Figure 1*.





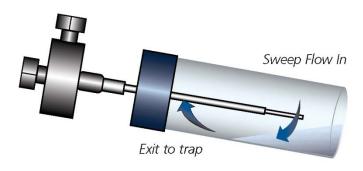


FIGURE 1 Patented dynamic headspace dual needle design.

### **RESULTS**

The flavor absorption of d-limonene and a-pinene by polymeric packaging materials, such as low-density polyethylene has been extensively investigated (K.S.M., Sheug, S. Min, and S. K. Sastry 2004). A small loss of flavor compounds will lead to serious changes in flavor quality (K.S.M., Sheug, S. Min, and S. K.

Sastry 2004). These two key favor compounds were examined in this study. The challenge is that d-limonene appears in high part per million levels and a-pinene can be in low parts per billion levels see *Figure 2*. The instrument operating conditions can be seen in *Tables 1&2* 

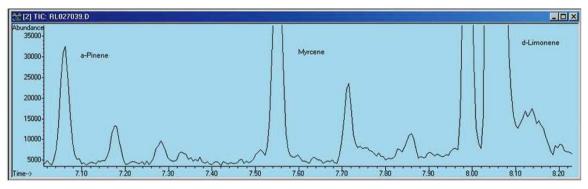


FIGURE 2 Orange juice carton NT

A container of orange juice obtained from the same distributor as in the research (K.S.M., Sheug, S. Min, and S. K. Sastry 2004) was used to examine 2 techniques. The first technique was the traditional loop fill and inject generally used for part per million levels of analysis. The second technique was achieved by simply switching the method in the software to the new dynamic sweep and trap technique that allows parts per billion levels of detection. A sealed sample was heated and mixed to reach equilibrium.

The sample vial was then placed onto the dual needle (a needle within a needle). Inert gas flows at a prescribed rate for a programmed period of time through one passage of the needle, displacing the headspace in the vial. The analytes were carried out of the vial through the second passage of the needle and concentrated onto a cool adsorbent trap. See Figure 1 The adsorbent trap is then heated and back-flushed with the GC carrier gas transporting the analytes to the GC column.



# TABLE 1 Instrument Operating Conditions

GC Cycle Time	25	
Constant Heat Mode	enable	
Vial Type	20	
Pressure Check	cont	
Sample Vial Temp	45	
Sample Equilibration Time	15	
Mixing	On	
Mixing Speed	med	
Post Mix Stabilization Time	10	
Line Temp	155	
Valve / Loop Temp	150	
Standby Flow	On 40	
Economy Mode	program	
Sample Mode	2N Trap	

TABLE 2
Instrument Operating
Conditions

Trap	Tenax
Trap Ready Temp	40
Pretrap Ready Temp	25
Sweep Flow Rate	20
Sweep Time	2
Trap Dry Sweep Temp	25
Dry Sweep Flow Rate	80
Dry Sweep Time	1
Trap Desorb Preheat Temp	210
Trap Desorb Temp	220
Trap Desorb Time	2
Trap Bake Temp	220
Pretrap Bake Temp	25
Bake Flow Rate	85
Bake Time	10

The same needle was used to perform traditional static headspace injections giving the analyst the ability to make both static and dynamic injections within the same sample schedule. The HS9000 not only delivers improved sensitivity with the dynamic sweeping ability but also delivers the proven technology

of static equilibrium headspace analysis all in the same instrument. The two main favor compounds of interest in this study (d-limonene and a-pinene) were easily examined by the automated techniques see *Figures 3 & 4*.

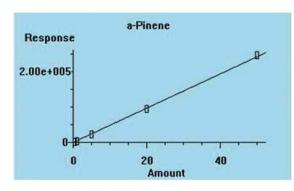


FIGURE 3 0.5-50 ppb Cal curve

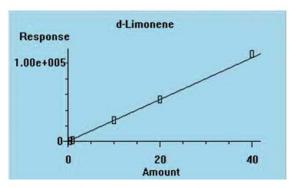


FIGURE 4 0.5-40 ppb Cal curve

The high precision and linearity examining the volatiles present in a complex mixture at such a wide range of levels is extraordinary Tables 3& 4. Method optimization can be completely automated by testing the variables such as the equilibration times, pressures, sweep time, flow time and temperatures with just a change in the

method software The analysis of product adulteration, impurity contamination, and off-odor characteristics can now be precisely examine with never before simplicity and ease of use.

## TABLE 3

Compound	0.5ppb	1ppb	5ppb	20ppb	50ppb	Avg	% RSD
a-Pinene	4.940	4.697	4.769	4.728	4.961	4.819 E3	2.55

# **TABLE 4**

Compound	0.5ppb	1ppb	10ppb	20ppb	40ppb	Avg	% RSD
d-Limonene	2.716	2.304	2.688	2.705	2.789	2.640 E3	7.27

### CONCLUSION

A new simple, automated, static/dynamic headspace gas chromatography technique can be used to determine the absorption of flavor absorption into the packaging material. The new dual needle trapping technique provides a robust sample introduction method at unprecedented low levels of detection of favors in packaging. Flavor compounds in food exist in trace levels that are very important concern to the food/package manufacturing process. Now researchers have a tool capable of measuring sub ppb levels with ease of use, accuracy and precision.

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